



**CHEMSEA**  
CHEMICAL MUNITIONS  
SEARCH & ASSESSMENT

# ANALYSIS OF CHEMICAL WARFARE AGENT RELATED CHEMICALS IN *IN VIVO* EXPOSED MUSSELS

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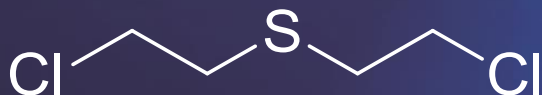
Part-financed by  
the European Union  
(European Regional  
Development Fund)

# Studied dumping areas

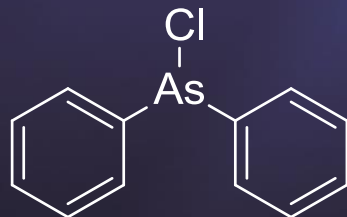


# Major chemicals

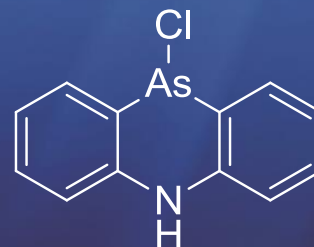
and amounts dumped in Bornholm



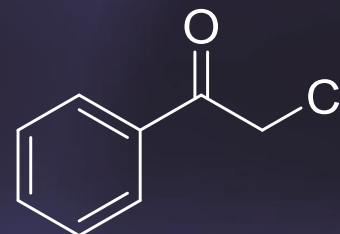
Sulphur mustard (H)  
7 000 tons



Clark I (DA)  
~1 000 tons



Adamsite (DM)  
1 500 tons



$\alpha$ -Chloroacetophenone (CN)  
500 tons

- Samples from mussel exposure experiment (FOI/SYKE/AWI)
  - The main aim was to evaluate biological responses in blue mussels (*Mytilus edulis* L.) induced by chemical warfare agents (CWA) mixtures at environmentally relevant concentrations
  - Mussels were exposed to 11 different mixtures containing the arsenic containing warfare agents Diphenylarsinechloride (Clark I, DA), 10-Chloro-5-hydrophenarsazine (Adamsite, DM) and the tear gas  $\alpha$ -Chloroacetophenone
  - Different concentrations (low, medium, high) were selected for each compound ranging from 1.25 to 50  $\mu\text{g/L}$  based on the assumed toxicity of each chemical
  - Each mixture group consisted of 56 mussels that were distributed among eight glass aquaria (12L; 7 mussels per aquarium)
  - A separate exposure was conducted with bis(2-chloroethyl)sulfide (mustard agent) only

<b>Experimental design for the testing of:</b>									
<b>one species (cultivated blue mussels, 2nd year, 2.5-3.5 cm)</b>									
<b>Present example is a setup for:</b>									
11 mixtures (+ 2 for the middle point) = 13 treatments									
<b>32 aquaria, 6 (+ 1 as back up) individuals per aquaria</b>									
<b>Sampling of 0-Group before start of experiment</b>									
<b>Daily water exchange, feeding once a week</b>									

Totally 56 mussels/mixture group  
Whole experiment lasted for 6 weeks

[illegible]



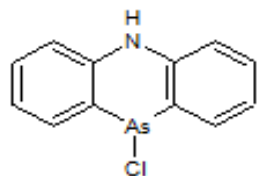
# Adamsite, Clark, $\alpha$ -Chloroacetophenone and Mustard Gas added at environmentally relevant concentrations



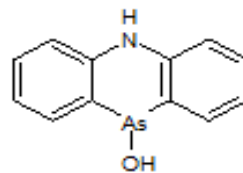
- No published data of metabolism or detoxification rates of CWAs in mussel
- For the evaluation of the risks of dumped CWAs, chemicals which pose the highest realistic risk to marine biota are:
  - Sulfurd mustard (H)
  - Adamsite (DM)
  - Clark I (DA)
  - Triphenyl arsine (TPA)
- Probable metabolic reactions are hydrolysis and oxidation

## Hydrolysis

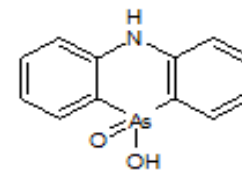
## Oxidation



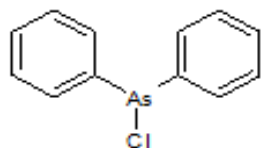
Adamisite (DM)



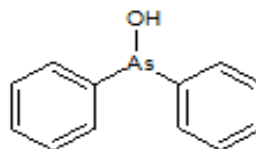
Phenoarsazin-10(5H)-ol



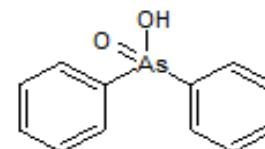
5,10-Dihydrophenoarsazin-10-ol 10-oxide (DMox)



Clark I (DA)



Diphenylarsinous acid



Diphenylarsinic acid (DAox)

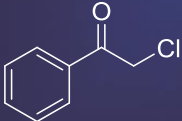
$\alpha$ -Chloroacetophenone has been reported to undergo hydrolysis slowly (Missianen et al 2010).



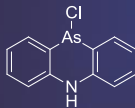
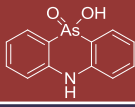
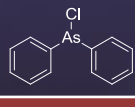
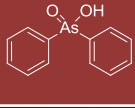
# Adamsite and Clark related products in water from mussel exposure experiments

- The measured concentrations of DMox and DAox (after oxidation) were generally lower than the nominal concentrations.
- There were no significant differences in concentration before and after the exposure period; however, the variation in the data was large.
- The reasons for the large variation could be related to inhomogeneity of DMox and DAox in the water (i.e. the compounds stick to glass or stay at the surface).
- It should also be noticed that the samples have been frozen for several months before analysis.

## ■ Analytes and analysis techniques – GC-MS/MS

#	Chemical (acronym) CAS	Structure	Description	GC-based			LC-based
				Intact	HFBI	TiCl <sub>3</sub>	Intact
	CN α-chloroacetophenone 532-27-4		Dumped CW agent	<i>Mussels (12)</i>			

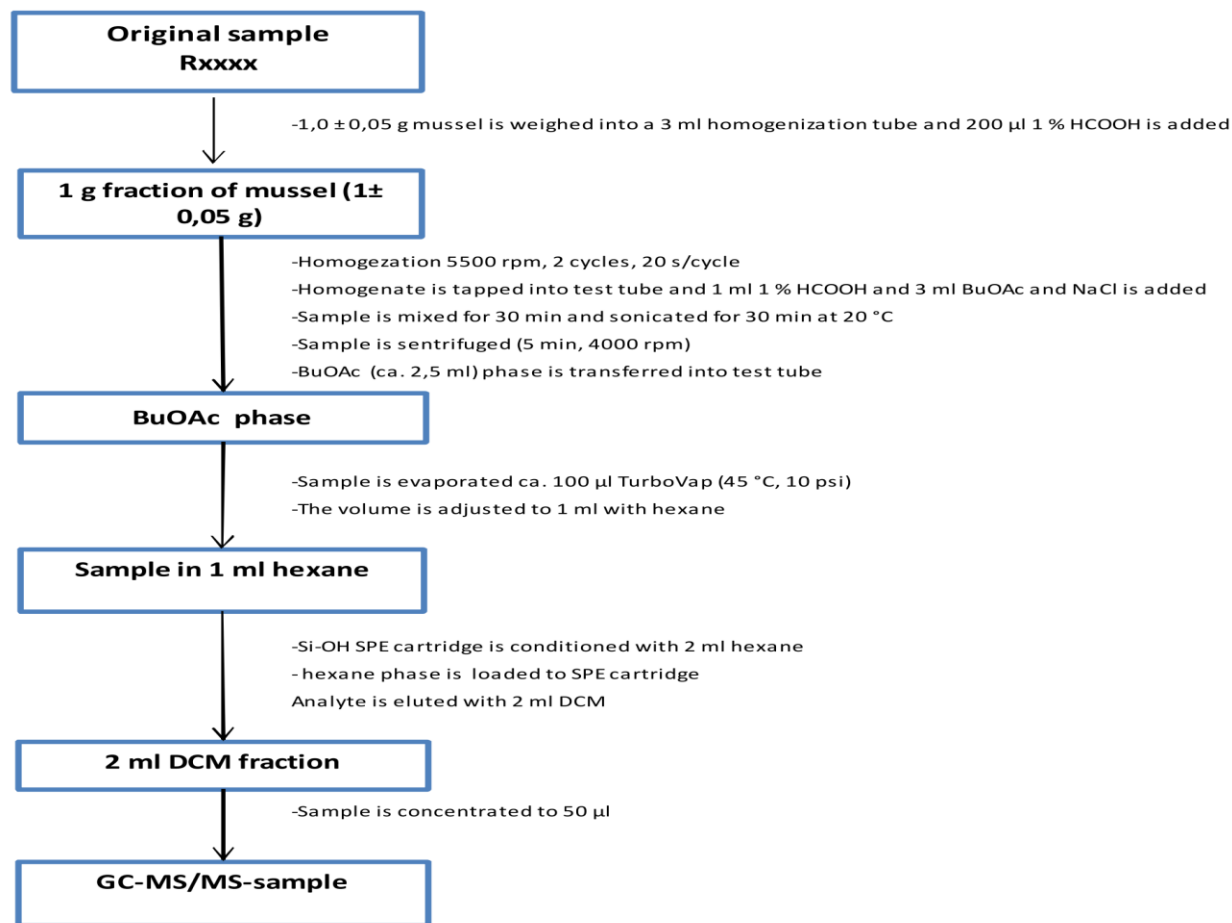
## ■ Analytes and analysis techniques – LC-MS/MS

#	Chemical (acronym) CAS	Structure	Description	GC-based			LC-based
				Intact	HFBI	TiCl <sub>3</sub>	Intact
2	Adamsite (DM) 578-94-9		Dumped CW agent	<i>Not analysed as such</i>			
20	5,10-Dihydrophenoarsazin-10-ol 4733-19-1		Oxidation product of <b>2</b> and all of its degradation products (either natural or with H <sub>2</sub> O <sub>2</sub> )				Exposed mussels (17)
3	Clark I (DA) 712-48-1		Dumped CW agent. Also component in dumped arsine oil.	<i>Not analysed as such</i>			
30	Diphenylarsinic acid 4656-80-8		Oxidation product of <b>3</b> (either natural or with H <sub>2</sub> O <sub>2</sub> )				Exposed mussels (17)

# Sample preparation for GC-MS/MS biomarkers of $\alpha$ -Chloroacetophenone (CN)

- Mussel (CN)
  - Homogenization of 1.0 g of mussel + 200  $\mu$ l 1 % HCOOH
  - Addition of 1 % HCOOH, BuOAc and NaCl
  - Mixing, sonication and centrifugation
  - Concentration of BuOAc phase and volume adjusting to 1 ml with hexane
  - SPE clean-up
  - DCM fraction
  - Concentration

# Sample preparation for GC-MS/MS biomarkers of $\alpha$ -Chloroacetophenone (CN)





# Sample preparation for LC-MS/MS biomarkers of Adamsite, Clark

- Mussel (DM-ox,DA-ox)
  - Homogenization of 3.0 g of cod muscle/mussel meat + 1 ml of water
  - Addition of water, ACN and buffered salts
  - Mixing and centrifugation
  - ACN layer, addition of hexane to remove excess fat
  - Shaking
  - Discarding the hexane layer and separating ACN layer
  - Concentration of the sample
  - Dilution of sample with water and filtration
  - Addition of 25 µl ISTD (DMMP)

- GC-MS/MS analysis –CN
  - Triple quadrupole mass spectrometer with GC and autosampler
  - Capillary column DB-5ms (Agilent, 30 m x 0.25 mm i.d., 0.25 µm film)
  - The column temperature programme: 40°C (isothermal time 1 min) to 320 °C, 20 °C/min
  - Transfer line temperature 325 °C
  - Splitless injection (splitless time 1 min, 1 µl)
  - Injector temperature 250°C
  - EI (70 eV)
  - Selected reaction monitoring (SRM)
  - LOQ 0.2 ng/g for CN

- LC-MS/MS analysis - DM-ox and DA-ox
  - Triple quadrupole mass spectrometer with two LC pumps and autosampler
  - Waters XBridge BEH C18 column (2.1 x 100 mm, 2.5  $\mu$ m) + Pre-Column
  - Column temperature 30°C
  - Flow-rate 0.4 ml/ml
  - Injection volume 5 $\mu$ l
  - Eluent (A) 0.1% formic acid in water
  - Eluent (B) 0.1% formic acid in MeOH
  - APCI in positive mode by SRM
  - Mussels: LOQ 10 ng/ml for DMox and Daox

# Results of mussel samples after Adamsite *in vivo* exposure

Code	DMox (ng/g, ppb)	DM in exposure mixture (ng/ml)
Blank 1	NF	No DM added
Blank 2	NF	No DM added
Control	NF	No DM added
Solvent Control	NF	No DM added
N1	<b>77</b>	<b>2.5</b>
N2	<b>339</b>	<b>10</b>
N3	<b>128</b>	<b>2.5</b>
N4	<b>358</b>	<b>10</b>
N6	<b>586</b>	<b>10</b>
N5	<b>280</b>	<b>2.5</b>
N7	<b>159</b>	<b>2.5</b>
N8	<b>394</b>	<b>5.0</b>
N9	<b>366</b>	<b>10</b>
N10	<b>209</b>	<b>5</b>
N11	<b>136</b>	<b>5</b>
N12	<b>99</b>	<b>5</b>
N13	<b>141</b>	<b>5</b>
Quality blank	NF	-
Blank mussel	NF	-

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-Quality blank	NF	-
-Blank mussel	NF	-



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N8	394	5.0
N9	366	10
N10	209	5
N11	136	5
N12	99	5
N13	141	5
-Quality blank	NF	-
-Blank mussel	NF	-

# Results of mussel samples after Clark *in vivo* exposure

Code	DAox (ng/g, ppb)	DA in exposure mixture (ng/ml)
Blank sample	NF	No DA added
Blank sample	NF	No DA added
Control	NF	No DA added
Solvent control	NF	No DA added
N1	<b>127</b>	<b>1.25</b>
N2	<b>62</b>	<b>1.25</b>
N3	<b>569</b>	<b>5.0</b>
N4	<b>337</b>	<b>5.0</b>
N6	<b>91</b>	<b>1.25</b>
N5	<b>90</b>	<b>1.25</b>
N7	<b>724</b>	<b>5.0</b>
N8	<b>584</b>	<b>5.0</b>
N9	<b>77</b>	<b>2.5</b>
N10	<b>98</b>	<b>2.5</b>
N11	<b>62</b>	<b>2.5</b>
N12	<b>133</b>	<b>2.5</b>
N13	<b>226</b>	<b>2.5</b>
Quality blank	NF	-
Blank mussel	NF	-

# Results of mussel samples after Clark *in vivo* exposure

Code	DAox (ng/g, ppb)	DA in exposure mixture (ng/ml)
Blank sample	NF	No DA added
Blank sample	NF	No DA added
Control	NF	No DA added
Solvent control	NF	No DA added
N1	<b>127</b>	<b>1.25</b>
N2	<b>62</b>	<b>1.25</b>
N3	<b>569</b>	<b>5.0</b>
N4	<b>337</b>	<b>5.0</b>
N6	<b>91</b>	<b>1.25</b>
N5	<b>90</b>	<b>1.25</b>
N7	<b>724</b>	<b>5.0</b>
N8	<b>584</b>	<b>5.0</b>
N9	<b>77</b>	<b>2.5</b>
N10	<b>98</b>	<b>2.5</b>
N11	<b>62</b>	<b>2.5</b>
N12	<b>133</b>	<b>2.5</b>
N13	<b>226</b>	<b>2.5</b>
Quality blank	NF	-
Blank mussel	NF	-

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Blank sample	NF	No DA added
Blank sample	NF	No DA added
Control	NF	No DA added
Solvent control	NF	No DA added
N1	127	1.25
N2	62	1.25
N3	569	5.0
N4	337	5.0
N6	91	1.25
N5	90	1.25
N7	724	5.0
N8	584	5.0
N9	77	2.5
N10	98	2.5
N11	62	2.5
N12	133	2.5
N13	226	2.5
Quality blank	NF	-
Blank mussel	NF	-

# Results of mussel samples after Clark *in vivo* exposure

Code	DAox (ng/g, ppb)	DA in exposure mixture (ng/ml)
Blank sample	NF	No DA added
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Solvent control	NF	No DA added
N1	<b>127</b>	<b>1.25</b>
N2	<b>62</b>	<b>1.25</b>
N3	<b>569</b>	<b>5.0</b>
N4	<b>337</b>	<b>5.0</b>
N6	<b>91</b>	<b>1.25</b>
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N9	<b>77</b>	<b>2.5</b>
N10	<b>98</b>	<b>2.5</b>
N11	<b>62</b>	<b>2.5</b>
N12	<b>133</b>	<b>2.5</b>
N13	<b>226</b>	<b>2.5</b>
Quality blank	NF	-
Blank mussel	NF	-



# Results of mussel samples after CN *in vivo* exposure

Code	CN (ng/g, ppb)	CN in exposure mixture (ng/ml)
Solvent control	NF	No CN added
N1	NF	5
N2	1.3	50
N4	1.2	5
N5	NF	50
N6	1.3	5
N7	NF	5
N8	1.6	15.8
N9	NF	15.8
N10	NF	50
N11	NF	16
N12	NF	15.8
N13	1.7	15.8
Quality blank	NF	-
Blank mussel	NF	-

- Improve sample preparation methods for analysis
- Need for internal standards, stable isotope labelled reference chemicals appreciated
- New reference chemicals to be synthesized e.g. Glutathione conjugates
- Toxicity studies
- Metabolism in mussels

- Real proof for the source of ecotoxical effects can only be found after chemical analysis of degradation products/metabolites in mussel samples
- EU identification criteria followed
- High concentrations of oxidized DM and DA were measured the soft tissue from in-vivo exposed mussels